

# The Antiviral Activities of Artemisinin and Artesunate

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Traditional Chinese medicine commands a unique position among all traditional medicines because of its 5000 years of history. Our own interest in natural products from traditional Chinese medicine was triggered in the 1990s, by artemisinin-type sesquiterpene lactones from *Artemisia annua* L. As demonstrated in recent years, this class of compounds has activity against malaria, cancer cells, and schistosomiasis. Interestingly, the bioactivity of artemisinin and its semisynthetic derivative artesunate is even broader and includes the inhibition of certain viruses, such as human cytomegalovirus and other members of the *Herpesviridae* family (e.g., herpes simplex virus type 1 and Epstein-Barr virus), hepatitis B virus, hepatitis C virus, and bovine viral diarrhea virus. Analysis of the complete profile of the pharmacological activities and molecular modes of action of artemisinin and artesunate and their performance in clinical trials will further elucidate the full antimicrobial potential of these versatile pharmacological tools from nature.

Artemisinin is a natural product derived from the Chinese herb *Artemisia annua* (figure 1). During the Vietnam War, Ho Chi Minh asked Mao Zedong for help, because more North Vietnamese soldiers were dying from malaria than from armed conflicts. The Chinese government launched a program to find new antimalarial drugs. As a result, Tu Youyou, a Chinese scientist from the Chinese Academy of Traditional Chinese Medicine (Beijing, China), identified artemisinin as the active compound of *A. annua* in 1972 [1]. Its overwhelming antimalarial activity was demonstrated in numerous clinical studies by Chinese and Western scientists. Despite this success, the true potential of artemisinin was underestimated in the Western world for many years

[2]. In the meantime, the World Health Organization officially recommends artemisinin and its derivatives, such as artesunate and artemether, for the treatment of malaria, particularly as a part of combination therapies with other antimalarial drugs.

Artemisinin, artesunate, and additional derivatives are the most promising candidate compounds to ease the worldwide malaria burden. The high safety and tolerability profile of these drugs adds to their attractiveness [3]. This group of compounds is also active against cancer cells and schistosomiasis [4–10]. The focus of the present review is the antiviral activity of artemisinin and artesunate.

Although some authors claim that the heme-mediated decomposition of the endoperoxide bridge and production of carbon-centered free radicals is necessary for antimalarial activity [11], other data indicate that the biological activity of artemisinin-like drugs does not correlate with their chemical reactivity [12]. Computer-assisted models for the calculation of quantitative structure-activity relationships have been developed to address these contradictory results [13–15].

Peak plasma concentrations of 391–588  $\mu\text{g/L}$  have

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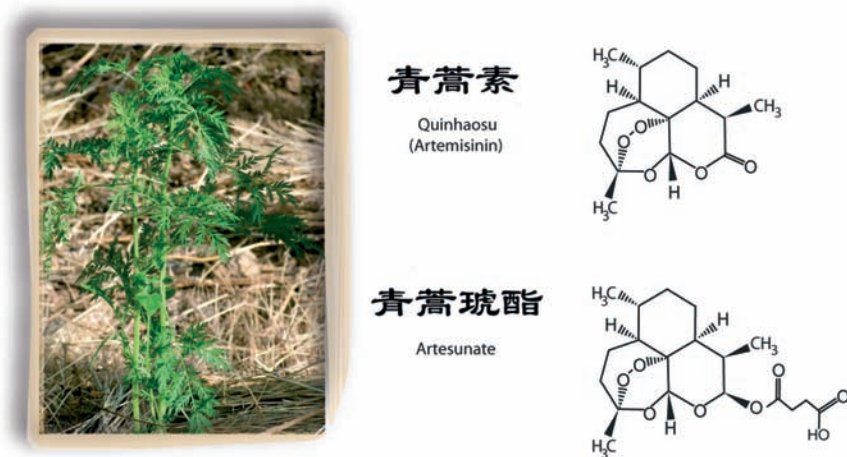
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**Figure 1.** Cultivar of the Chinese herb *Artemisia annua* L. (qin hao or sweet wormwood) and its active principle artemisinin (qin haosu). Artesunate represents a semisynthetic derivative of artemisinin that has improved water solubility.

been reported for orally administered artemisinin (500 mg, single dose) [16, 17], and peak plasma concentrations of 2640  $\mu\text{g/L}$  and 2020  $\mu\text{g/L}$  have been reported for intravenous artesunate (2 mg/kg) and for its active metabolite dihydroartemisinin, respectively [18, 19]. The oral administration of artesunate (100 mg) leads to a plasma-elimination half-life of 39–95 min for dihydroartemisinin [20–26].

The metabolism of artemisinin in human liver microsomes is primarily mediated by cytochrome P-450 monooxygenase enzyme (CYP) 2B6, with a secondary contribution by CYP3A4 in individuals with low CYP2B6 expression. The contribution of CYP2A6 to artemisinin metabolism is likely of minor importance [27]. There is a large body of evidence suggesting that artemisinin influences the CYP activity, which could result in drug-drug interactions [28]. An induction of activity by artemisinin was reported for CYP2A5, CYP2A6, CYP2B1, CYP2B6, CYP2B10, CYP2C19, and CYP3A4 [29–34]. In addition, artemisinin activates the constitutive androstane receptor and pregnane X receptor [33, 34], which explains the upregulation of CYP2B6 and CYP3A4. The data regarding CYP1A2 are contradictory [35–38], whereas artemisinin inhibits CYP2D6 [37]. Artemisinin leads to an autoinduction of drug metabolism, which reduces its own bioavailability [39].

In various clinical studies, artemisinin has been administered alone or in combination with other antimalarial drugs in dosages of up to 500 mg per day. As reviewed elsewhere, clinical trials of artesunate monotherapy used dosages of 1–8 mg/kg intravenously or 600–1200 mg per day orally for 5 days [40]. In combination therapies, 4–25 mg/kg intravenously or 200–800 mg per day orally for 3 days have been used [40].

Artemisinin derivatives are tolerated well by patients [41, 42]. Mild and reversible hematological and electrocardiographic abnormalities, such as neutropenia and first-degree heart block,

have been observed infrequently. Neurotoxic effects have been repeatedly reported in experiments with mice, rats, and dogs, as reviewed elsewhere [43]. Affected areas in the brain stem are the reticular system with regard to autonomic control, the vestibular system, the auditory system (trapezoid nucleus), and the red nucleus, which is important for coordination [44–51]. A longer exposure time to a lower peak blood concentration of an artemisinin derivative is more neurotoxic than a shorter duration of exposure and a higher peak blood concentration [52]. These animal experiments gave rise to concerns about the safety of artemisinin and its derivatives in humans. A clinical safety review of 108 clinical studies that enrolled 9241 malaria patients provided ample evidence that artemisinins are safe and without serious adverse effects or significant severe toxicity, including neurotoxicity [41]. Ataxia, slurred speech, and hearing loss have been reported in few patients treated with artemisinin [53]. Although the artemisinin derivative artesunate seems to be without toxicity, van Hensbroek et al. [54] observed delayed coma recovery times in Gambian children with malaria who were treated with intramuscular artemether versus intravenous quinine. Because of these conflicting results, Stepniewska et al. [55] performed a meta-analysis of 7 studies involving 1919 patients with malaria. Applying a uniform coma recovery time definition, no significant difference in coma recovery time was found between patients treated with artemether and quinine. Additionally, no statistically significant difference was observed with regard to neurological sequelae. In a recent study by Dondorp et al. [56], patients with malaria who were treated with artesunate were compared with patients who were treated with quinine. The authors did not find significant differences in terms of neurotoxic symptoms (i.e., times to speak, eat, and sit) between treatment groups. Neurological sequelae did not occur after treatment. Interestingly, patients with ma-

laria who developed late onset hypoglycemia had a higher incidence of death than did patients treated with artesunate who did not have hypoglycemia. This may be an issue that deserves additional investigation.

## ACTIVITY AGAINST HUMAN CYTOMEGALOVIRUS (HCMV)

Chinese scientists provided the first hint that artemisinin might have antiviral activity [57]. Indeed, artesunate inhibits the *in vitro* replication of HCMV (HCMV AD169 and other strains; table 1) and herpes simplex virus type 1 (HSV-1) [59]. With regard to artesunate's potential inhibition of HCMV, it was important to demonstrate that viruses with a variety of phenotypes (i.e., low-passage clinical isolates, drug-resistant mutants, laboratory strains, and recombinant virus clones) were all highly sensitive to artesunate. A possible mechanism was suggested by the finding that artesunate inhibited central regulatory processes of HCMV-infected cells (such as activation pathways dependent on NF- $\kappa$ B or Sp1), thus interfering with critical host-cell-type and metabolism requirements for HCMV replication.

HCMV is a major cause of disease in immunocompromised individuals, including patients with AIDS and transplant recipients, and it is a common cause of congenital infection leading to developmental abnormalities and hearing loss [62]. All currently available anticytomegaloviral drugs, including ganciclovir, foscarnet, and cidofovir, target the viral DNA polymerase. The use of these drugs is limited by toxicity, low oral bioavailability (with the exception of the oral prodrug valganciclovir), teratogenicity, and drug resistance. These limitations,

along with the repeated and prolonged courses of therapy often required for the treatment of HCMV infection in transplant recipients, create an increasing need for new antiviral drugs, particularly for drugs that exhibit low levels of toxicity and activity against HCMV variants that are resistant to conventional drugs [59].

The replication of HCMV is tightly coregulated with cellular activation pathways mediated by the direct or indirect interaction with cellular DNA-binding factors, such as NF- $\kappa$ B and Sp1 [63, 64]. These factors provide major determinants of the virus-host cell interaction. For both NF- $\kappa$ B and Sp1, a reduction in HCMV-induced protein synthesis and a reduction in the DNA binding activity of NF- $\kappa$ B and Sp1 were observed with artesunate treatment [59]. The inhibitory activity towards NF- $\kappa$ B is not specific for artesunate alone; it has also been demonstrated for other sesquiterpene lactones [65, 66]. The efficiency of HCMV replication is closely connected with NF- $\kappa$ B and Sp1 activation pathways and other involved factors, such as the cellular signaling kinase phosphoinositol 3-kinase [67]. Phosphoinositol 3-kinase is required for the activation of NF- $\kappa$ B and Sp1 in infected fibroblasts. Interestingly, the phosphorylation of downstream effectors of phosphoinositol 3-kinase, such as the protein kinases Akt and p70S6K, is also inhibited by artesunate [59] (figure 2). Moreover, there are several examples that chemical compounds interfering with activation pathways of cellular transcription factors (e.g., the signal transduction pathway that includes mitogen-activated protein kinase p38) inhibit HCMV replication. It is noteworthy that the HCMV immediate-early promoter enhancer (in addition to other viral promoters) contains binding sites for both Sp1 and

**Table 1. Sensitivity of herpesviruses to artesunate.**

Sensitivity test	Herpesvirus type	Herpesvirus subfamily	Strain or isolate	Type of analysis	Inhibition at 15 $\mu$ M, %	IC <sub>50</sub> , $\mu$ M	Reference
1	HCMV	$\beta$	AD169	In vitro <sup>a</sup>	...	3.9 $\pm$ 0.6	[58]
2	HCMV	$\beta$	AD169	In vitro <sup>a</sup>	81	5.8 $\pm$ 0.4	[59]
3	HCMV	$\beta$	Towne	In vitro <sup>a</sup>	>99	...	[59]
4	HCMV	$\beta$	Clinical isolates	In vitro <sup>a</sup>	82	...	[59]
5	HCMV	$\beta$	Clinical isolates	In vitro <sup>a</sup>	69	...	[59]
6	HCMV	$\beta$	Ganciclovir resistant mutant	In vitro <sup>a</sup>	...	6.9 $\pm$ 0.2	[59]
7	HCMV	$\beta$	Multidrug resistant mutant	Clinical trial <sup>b</sup>	...	...	[60]
8	RCMV	$\beta$	Maastricht	In vitro <sup>a</sup>	38	...	[58]
9	RCMV	$\beta$	Maastricht	In vivo model <sup>c</sup>	...	...	[58]
10	MCMV	$\beta$	Smith	In vitro <sup>a</sup>	15	...	M.M., unpublished data
11	HHV-6A	$\beta$	U1102	In vitro <sup>d</sup>	76	3.80 $\pm$ 1.06	M.M., unpublished data
12	HSV-1	$\alpha$	Clinical isolate	In vitro <sup>a</sup>	83	...	[59]
13	EBV	$\gamma$	B95-8	In vitro <sup>d</sup>	63	7.21 $\pm$ 2.25	M.M., unpublished data

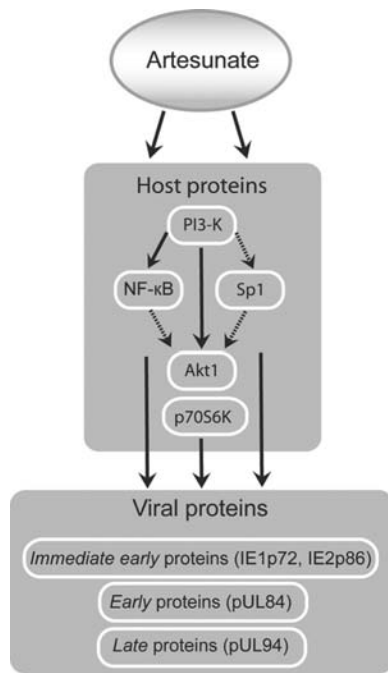
**NOTE.** EBV, Epstein-Barr virus; HCMV, human cytomegalovirus; HHV-6A, human herpesvirus 6A; HSV-1, herpes simplex virus 1; MCMV, murine cytomegalovirus; RCMV, rat cytomegalovirus.

<sup>a</sup> Virus replication analyzed in infected primary fibroblasts (green fluorescent protein-based reporter assays and plaque reduction assays) [61].

<sup>b</sup> Clinical trial with HCMV-infected patients after hematopoietic stem cell transplantation (quantitative PCR and antigenemia assays for viral load in blood specimens) [60].

<sup>c</sup> Virus replication analyzed in immunosuppressed rats (quantitative PCR and plaque reduction assays of salivary gland specimens) [58].

<sup>d</sup> Virus replication analyzed in infected immortalized lymphocytes (quantification of immunofluorescence stainings).



**Figure 2.** Hypothetical molecular mechanism of artesunate against human cytomegalovirus.

NF- $\kappa$ B and, therefore, is responsive to both factors [68, 69]. Reduction of IE2p86 expression critically limits viral replication, because IE2p86 is essential for the initiation of subsequent regulatory steps [70]. On the other hand, NF- $\kappa$ B is a major factor in cellular defense pathways (e.g., IFN type 1-induced antiviral effects) and may also have a negative impact on viral productivity and the course of infection [71]. Thus, the activation pathways that involve Sp1 and NF- $\kappa$ B are important factors in the initial onset of the viral replication cycle, as well as in later steps in virus-cell interaction, and are, therefore, crucial for the antiviral action of artesunate.

Because of this background, it was interesting to analyze whether artesunate was active against drug-resistant HCMV, as well. Indeed, ganciclovir-resistant mutants (i.e., the laboratory mutant AD169-GFP314, which carries the resistance-conferring mutation UL97 [M460I], or ganciclovir-resistant clinical isolates) were inhibited with similar efficacy as the drug-sensitive parental virus (AD169-GFP) [61]. It is obvious from the results of these experiments involving ganciclovir-resistant HCMV that the putative inhibitory mechanisms of artesunate must be different than the mechanism of conventional DNA polymerase-inhibiting drugs.

The anticytomegaloviral activity of artesunate is not restricted to HCMVs but also includes animal CMVs—in particular, rat CMV [58]. An important finding was that increased intracellular iron concentrations enhance artesunate's anticytomegaloviral activity. This iron-enhanced effectiveness was demonstrated by several observations, and the following are

some promising features of the anticytomegaloviral activity of artesunate. First, treatment of CMV-infected fibroblasts with artesunate plus ferrous iron (Ferrosanol; Monheim) and/or soluble Transferrin resulted in enhanced suppression of viral replication. Because Ferrosanol is a clinically approved formulation, this drug could potentially be safely combined with artesunate in clinical practice. Second, the antiviral activity of artesunate is additive in combination with conventional drugs, such as ganciclovir, cidofovir, and foscarnet. A combination of drugs with different modes of action may delay the development of drug resistance. Third, the antiviral activity of artesunate against CMV was also demonstrated in vivo using the rat CMV model. Importantly, the first successful clinical use of artesunate for the treatment of HCMV in a patient who developed drug-resistant infection during preemptive antiviral therapy after stem cell transplantation has been described [60]. In this case, artesunate proved to be a highly effective and well-tolerated inhibitor of HCMV replication, which suggests the need for additional clinical evaluation of its role in the treatment of HCMV infection.

## BROAD-SPECTRUM ACTIVITY AGAINST HERPES VIRUSES

The antiviral activity of artesunate is not restricted to distinct viral laboratory strains; artesunate is also effective against clinical isolates of HCMV and mutants with resistance against conventional antiviral drugs, such as ganciclovir and cidofovir (M. Leis and M.M., unpublished data). Novel data show that other herpesviruses from all subfamilies ( $\alpha$ ,  $\beta$ , and  $\gamma$ ) are also sensitive to artesunate—namely, Epstein-Barr virus, herpes simplex virus 1, and human herpes 6A (table 1; M.M., unpublished data) [58, 59]. This finding suggests that artesunate has broad activity against herpesviruses. The herpesviruses that have been analyzed thus far have all demonstrated similar sensitivity to artesunate ( $IC_{50}$ ,  $<10 \mu M$ ). Some of the analyzed herpesviruses show different sensitivities to artesunate and artemisinin; artemisinin is inactive against human herpesvirus 6A [72] and has poor activity against HCMV [58]. This indicates that the semisynthetic drug artesunate has more antiherpesviral potency than does its natural parental drug, artemisinin.

## ACTIVITY AGAINST HEPATITIS B VIRUS (HBV)

The family *Hepadnaviridae* includes a group of highly species-specific viruses that all have a virus-encoded DNA polymerase with reverse-transcriptase activity [73–75]. One member of this family, human HBV, is characterized by a high level of hepatotropism. This virus belongs to the genus *Orthohepadnavirus* and is not cytopathic itself, although it may cause acute fulminant hepatitis [76] or chronic liver disease, which may progress to cirrhosis and, eventually, hepatocellular carcinoma [77]. In spite of the availability of an effective and safe vaccine

against HBV, infection by this virus has remained a major worldwide health problem [78, 79]. Although several pharmacological strategies are currently being implemented to treat HBV-infected patients (i.e., the use of IFN and a nucleoside derivative, lamivudine), no effective antiviral therapy against HBV infection has yet been fully developed.

In a recent investigation [80], a panel of natural products derived from medicinal herbs used in traditional Chinese medicine has been assayed for anti-HBV activity. Among these products, artesunate displayed anti-HBV activity. HBV DNA release was inhibited at an  $IC_{50}$  of 0.5  $\mu$ M. Host cell viability was reduced at a concentration 40-times greater (20  $\mu$ M). Moreover, the treatment potential is enhanced by synergistic effects with lamivudine and by the absence of drug-induced toxicity in host cells. This is important in clinical practice because of frequent cases of infection by lamivudine-resistant HBV strains.

The concentration at which artesunate was active against HBV (>10  $\mu$ M) was similar to that previously reported for its activity against HCMV [59]. Interestingly, these levels are close to the drug concentrations achieved in the plasma of patients in whom this drug is used for anti-malarial treatment (~7  $\mu$ M) [81]. This result was similar to that reported elsewhere [82] for artesunate use in HepG2 2.2.15 cells.

### ACTIVITY AGAINST HEPATITIS C VIRUS (HCV) AND RELATED VIRUSES

The family *Flaviviridae* includes 3 genera: *Pestivirus* (e.g., bovine viral diarrhoea virus), *Flavivirus* (e.g., Japanese encephalitis virus), and *Hepacivirus* (e.g., HCV). Pathogens of the family *Flaviviridae* constitute a major cause of disease worldwide. Infection with HCV frequently causes chronic hepatitis, which may progress to cirrhosis and hepatocellular carcinoma [83]. The problem is aggravated by the absence of an efficient vaccine

against HCV and because the standard treatment (pegylated IFN- $\alpha$  and the purine nucleoside analogue ribavirin [ $1\beta$ -D-ribofuranosyl-1,2,4-triazole-3-carboxamide]), in addition to having adverse effects, is not effective in approximately one-half of infected patients [83]. Therefore, the search for more effective therapies is crucial. Because all members of the *Flaviviridae* share similarities in virion structure, genome organization, and replication machinery, some viruses, in particular bovine viral diarrhoea virus, have been used as in vitro models [84].

The pharmacological interest in artemisinin and its derivatives for the treatment of infections by these viruses is increased by the severe limitations of currently available antiviral therapy. Because the mechanisms of action of IFN- $\alpha$  [85, 86] and ribavirin [86, 87] against *Flaviviridae* viruses are probably different than the mechanisms of artemisinin [88], it was possible that a combination of these drugs would demonstrate additive effects; indeed, additive effects were observed by Romero et al. [89]. IFN binds to cell surface receptors and stimulates signal pathways that lead to the activation of cellular enzymes that repress viral replication [85], whereas ribavirin, in addition to its immunomodulatory properties, has direct antiviral activities that can be ascribed to several possible mechanisms. These mechanisms include the inhibition of the HCV RNA-dependent RNA polymerase NS5B and ribavirin's activity as an RNA mutagen, which enables it to impair viral replication [90]. Pae-shuyse et al. [91] reported that the antimalarial drug artemisinin inhibited HCV replicon replication in a dose-dependent manner in 2 HCV subgenomic replicon constructs at concentrations that did not affect Huh 5-2 host cells. Hemin, an iron donor, inhibits HCV replicon replication by inhibiting the viral polymerase [92]. The combination treatment of artemisinin and hemin had a pronounced synergistic antiviral activity without affecting host cells.

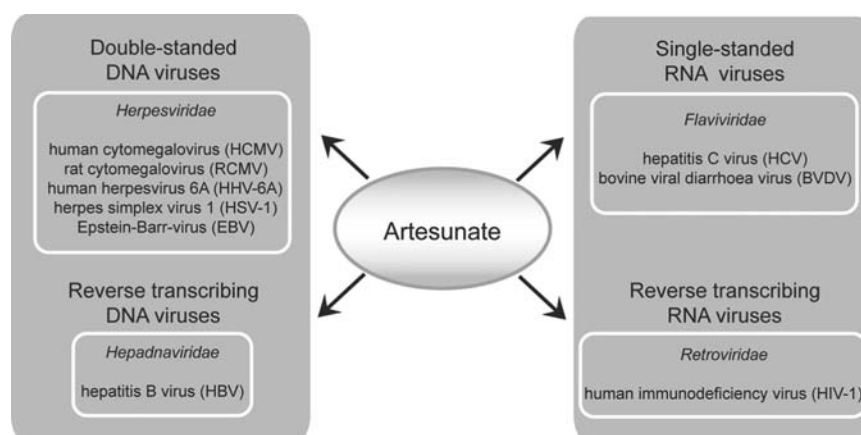


Figure 3. Spectrum of antiviral activity of artesunate

## ACTIVITY AGAINST HIV-1

Artesunate has been observed to have activity against HIV-1, as reported elsewhere [59]. Partial inhibition was demonstrated for 2 strains of HIV-1—the CCR5-tropic (M-tropic) HIV-1 strain Ba-L (in PM1 cells) and the CXCR4-tropic (T-tropic) HIV-1 strain NL4-3 (in Jurkat cells). The replication of both HIV-1 strains was partially inhibited by 600 nM artesunate throughout the analyzed time period of 10 days. Birku et al. [93] investigated the effect of artemisinin on the rate of clearance of *Plasmodium falciparum* in patients with or without HIV coinfection. Interestingly, Birku et al. [93] observed that HIV-infected patients showed a delayed clearance of *P. falciparum*, which suggested that the health of the host's immune system affects the activity of antimalarial drugs. No anti-HIV activity of artemisinin was reported in this investigation.

## CONCLUSIONS AND PERSPECTIVES

After being used in traditional Chinese medicine for 2 millennia, 1 of the “gems” of traditional Chinese medicine's treasure box has been rediscovered during recent years. Artemisinin is certainly one of the most promising natural products investigated in the past 2 decades. With regard to malaria, artemisinin has the potential to considerably contribute to a change in the desperate situation that the world is facing. Fortunately, the value of this compound is not limited to the treatment of malaria, and a wealth of studies have demonstrating the activity of artemisinin and its derivatives against cancer cells, schistosomiasis, and as reviewed here, various viral diseases (figure 3).

Ironically, in an age in which many scientists are searching for compounds with increased specificity to their molecular and cellular targets, awareness of artemisinin is increasing because of its multifunctionality. This class of compounds seems to have several targets that are important for different diseases. Conceptually, modern projects in molecular pharmacology aim to increase treatment efficacy and to decrease unwanted side effects by developing compounds that attack disease-related target molecules with high affinity. It is obvious that the natural evolution of pharmacologically active compounds in plants evolved in a different way. Natural products have evolved in plants as chemical weapons to protect against infections by bacteria, viruses, and other microorganisms. It is no surprise that multifunctional molecules might be more versatile and, therefore, more successful than monospecific molecules for protecting plants from environmental harm. In the case of artemisinin, it has been shown that it is active against various plant pathogenic fungi (i.e., *Gaeumannomyces graminis* var. *tritici*, *Rhizoctonia cerealis*, *Gerlachia nivalis*, and *Verticillium dahliae*) [94], which supports the role of artemisinin as a protective

agent for the plant. This view of chemical evolution in plants may fertilize current scientific concepts.

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## References

1. Tu Y. The development of new antimalarial drugs: qinghaosu and dihydro-qinghaosu. *Chin Med J (Engl)* **1999**; 112:976-7.
2. Attaran A, Barnes KI, Curtis C, et al. WHO, the Global Fund, and medical malpractice in malaria treatment. *Lancet* **2004**; 363:237-40.
3. Adjuk M, Agnamey P, Babiker A, et al. Artesunate combinations for treatment of malaria: meta-analysis. *Lancet* **2004**; 363:9-17.
4. Efferth T, Rücker G, Falkenberg M, et al. Detection of apoptosis in KG-1a leukemic cells treated with investigational drugs. *Arzneimittelforschung* **1996**; 46:196-200.
5. Efferth T, Dunstan H, Sauerbrey A, Miyachi H, Chitambar CR. The anti-malarial artesunate is also active against cancer. *Int J Oncol* **2001**; 18:767-73.
6. Efferth T, Sauerbrey A, Olbrich A, et al. Molecular modes of action of artesunate in tumor cell lines. *Mol Pharmacol* **2003**; 64:382-94.
7. Efferth T. Mechanistic perspectives for 1,2,4-trioxanes in anti-cancer therapy. *Drug Resist Updat* **2005**; 8:85-97.
8. Efferth T. Molecular pharmacology and pharmacogenomics of artemisinin and its derivatives in cancer cells. *Curr Drug Targets* **2006**; 7: 407-21.
9. Efferth T. Willmar Schwabe award 2006: antiplasmodial and antitumor activity of artemisinin—from bench to bedside. *Planta Med* **2007**; 73: 299-309.
10. Utzinger J, Xiao SH, Tanner M, Keiser J. Artemisinins for schistosomiasis and beyond. *Curr Opin Investig Drugs* **2007**; 8:105-16.
11. Meshnick SR. Artemisinin: mechanisms of action, resistance and toxicity. *Int J Parasitol* **2002**; 32:1655-60.
12. Haynes RK, Ho WY, Chan HW, et al. Highly antimalaria-active artemisinin derivatives: biological activity does not correlate with chemical reactivity. *Angew Chem Int Ed Engl* **2004**; 43:1381-5.
13. Avery MA, Alvim-Gaston M, Rodrigues CR, et al. Structure-activity relationships of the antimalarial agent artemisinin. 6. The development of predictive in vitro potency models using CoMFA and HQSAR methodologies. *J Med Chem* **2002**; 45:292-303.
14. Guha R, Jurs PC. Development of QSAR models to predict and interpret the biological activity of artemisinin analogues. *J Chem Inf Comput Sci* **2004**; 44:1440-9.
15. Cardoso FJ, de Figueiredo AF, da Silva Lobato M, et al. A study on antimalarial artemisinin derivatives using MEP maps and multivariate QSAR. *J Mol Model* **2008**; 14:39-48.
16. Zhao S. High-performance liquid chromatographic determination of artemisinin (qinghaosu) in human plasma and saliva. *Analyst* **1987**; 112:661-4.
17. Duc DD, de Vries PJ, Nguyen XK, et al. The pharmacokinetics of a single dose of artemisinin in healthy Vietnamese subjects. *Am J Trop Med Hyg* **1994**; 51:785-90.
18. Benakis A, Paris M, Plessas C, et al. Pharmacokinetics of sodium artesunate im and iv administration. *Am J Trop Med Hyg* **1993**; 49(Suppl):293.
19. Batty KT, Davis TME, Thu LT, Binh TQ, Anh TK, Ilett KF. Selective high-performance liquid chromatographic determination of artesunate and a- and b-dihydroartemisinin in patients with falciparum malaria. *J Chromatogr B Biomed Appl* **1996**; 677:345-50.

20. Teja-Isavadharm P, Watt G, Eamsila C, et al. Comparative pharmacokinetics and effect kinetics of orally administered artesunate in healthy volunteers and patients with uncomplicated falciparum malaria. *Am J Trop Med Hyg* **2001**;65:717–21.
21. Barradell LB, Fitton A. Artesunate: a review of its pharmacology and therapeutic efficacy in treatment of malaria. *Drugs* **1995**;50:714–41.
22. Batty KT, Thu LTA, Davis TME, et al. A pharmacokinetic and pharmacodynamic study of intravenous vs oral artesunate in uncomplicated falciparum malaria. *Br J Clin Pharmacol* **1998**;45:123–9.
23. Batty KT, Thu LTA, Ilett KF, et al. A pharmacokinetic and pharmacodynamic study of artesunate for vivax malaria. *Am J Trop Med Hyg* **1998**;59:823–7.
24. White NJ. Clinical pharmacokinetics and pharmacodynamics of artemisinin and derivatives. *Trans R Soc Trop Med Hyg* **1994**;88(Suppl 1):41–3.
25. Binh TQ, Ilett KF, Batty KT, et al. Oral bioavailability of dihydroartemisinin in Vietnamese volunteers and in patients with falciparum malaria. *Br J Clin Pharmacol* **2001**;51:541–6.
26. McGready R, Stepniewska K, Ward SA et al. Pharmacokinetics of dihydroartemisinin following oral artesunate treatment of pregnant women with acute uncomplicated falciparum malaria. *Eur J Clin Pharmacol* **2006**;62:367–71.
27. Svensson US, Ashton M. Identification of the human cytochrome P450 enzymes involved in the in vitro metabolism of artemisinin. *Br J Clin Pharmacol* **1999**;48:528–35.
28. Sukhija M, Medhi B, Pandhi P. Effects of artemisinin, artemether, arteether on the pharmacokinetics of carbamazepine. *Pharmacology* **2006**;76:110–6.
29. Mihara K, Svensson US, Tybring G, Hai TN, Bertilsson L, Ashton M. Stereospecific analysis of omeprazole supports artemisinin as a potent inducer of CYP2C19. *Fundam Clin Pharmacol* **1999**;13:671–5.
30. Giao PT, de Vries PJ. Pharmacokinetic interactions of antimalarial agents. *Clin Pharmacokinet* **2001**;40:343–73.
31. Li XQ, Björkman A, Andersson TB, Gustafsson LL, Masimirembwa CM. Identification of human cytochrome P(450)s that metabolise antiparasitic drugs and predictions of in vivo drug hepatic clearance from in vitro data. *Eur J Clin Pharmacol* **2003**;59:429–42.
32. Svensson US, Mäki-Jouppila M, Hoffmann KJ, Ashton M. Characterisation of the human liver in vitro metabolic pattern of artemisinin and auto-induction in the rat by use of nonlinear mixed effects modelling. *Biopharm Drug Dispos* **2003**;24:71–85.
33. Burk O, Arnold KA, Nussler AK, et al. Antimalarial artemisinin drugs induce cytochrome P450 and MDR1 expression by activation of xenosensors pregnane X receptor and constitutive androstane receptor. *Mol Pharmacol* **2005**;67:1954–65.
34. Simonsson US, Lindell M, Raffalli-Mathieu F, Lannerbro A, Honkakoski P, Lang MA. In vivo and mechanistic evidence of nuclear receptor CAR induction by artemisinin. *Eur J Clin Invest* **2006**;36:647–53.
35. Bapiro TE, Andersson TB, Otter C, Hasler JA, Masimirembwa CM. Cytochrome P450 1A1/2 induction by antiparasitic drugs: dose-dependent increase in ethoxyresorufin O-deethylase activity and mRNA caused by quinine, primaquine and albendazole in HepG2 cells. *Eur J Clin Pharmacol* **2002**;58:537–42.
36. Bapiro TE, Sayi J, Hasler JA, et al. Artemisinin and thiabendazole are potent inhibitors of cytochrome P450 1A2 (CYP1A2) activity in humans. *Eur J Clin Pharmacol* **2005**;61:755–61.
37. Asimus S, Elsherbin D, Hai TN, et al. Artemisinin antimalarials moderately affect cytochrome P450 enzyme activity in healthy subjects. *Fundam Clin Pharmacol* **2007**;21:307–16.
38. He F, Bi HC, Xie ZY, et al. Rapid determination of six metabolites from multiple cytochrome P450 probe substrates in human liver microsome by liquid chromatography/mass spectrometry: application to high-throughput inhibition screening of terpenoids. *Rapid Commun Mass Spectrom* **2007**;21:635–43.
39. Gordi T, Xie R, Huong NV, Huong DX, Karlsson MO, Ashton M. A semiphysiological pharmacokinetic model for artemisinin in healthy subjects incorporating autoinduction of metabolism and saturable first pass hepatic extraction. *Br J Clin Pharmacol* **2005**;59:189–98.
40. Barradell LB, Fitton A. Artesunate: a review of its pharmacology and therapeutic efficacy in treatment of malaria. *Drugs* **1995**;50:714–41.
41. Ribeiro IR, Olliaro P. Safety of artemisinin and its derivatives: a review of published and unpublished clinical trials. *Med Trop (Mars)* **1998**;58(Suppl 3):50–3.
42. Adjui M, Babiker A, Garner P, Olliaro P, Taylor W, White N. International Artemisinin Study Group. Artesunate combinations for treatment of malaria: meta-analysis. *Lancet* **2004**;363:9–17.
43. Toovey S. Are currently deployed artemisinins neurotoxic? *Toxicol Lett* **2006**;166:95–104.
44. Brewer TG, Grate SJ, Peggins JO, et al. Fatal neurotoxicity of arteether and artemether. *Am J Trop Med Hyg* **1994**;51:251–9.
45. Brewer TG, Peggins JO, Grate SJ, et al. Neurotoxicity in animals due to arteether and artemether. *Trans R Soc Trop Med Hyg* **1994**;88(Suppl 1):S33–6.
46. Genovese RF, Newman DB, Li Q, Peggins JO, Brewer TG. Dose-dependent brainstem neuropathology following repeated arteether administration in rats. *Brain Res Bull* **1998**;45:199–202.
47. Genovese RF, Newman DB, Petras JM, Brewer TG. Behavioral and neural toxicity of arteether in rats. *Pharmacol Biochem Behav* **1998**;60:449–58.
48. Kamchonwongpaisan S, McKeever P, Hossler P, Ziffer H, Meshnick SR. Artemisinin neurotoxicity: neuropathology in rats and mechanistic studies in vitro. *Am J Trop Med Hyg* **1997**;56:7–12.
49. Panossian LA, Garga NI, Pelletier D. Toxic brainstem encephalopathy after artemisinin treatment for breast cancer. *Ann Neurol* **2005**;58:812–3.
50. Petras JM, Kyle DE, Gettayacamin M, et al. Arteether: risks of 2-week administration in *Macaca mulatta*. *Am J Trop Med Hyg* **1997**;56:390–6.
51. Petras JM, Young GD, Bauman RA, et al. Arteether-induced brain injury in *Macaca mulatta*. I. The precerebellar nuclei: the lateral reticular nuclei, paramedian reticular nuclei, and perihypoglossal nuclei. *Anat Embryol* **2000**;201:383–97.
52. Li QC, Mog SR, Si YZ, Kyle DE, Gettayacamin M, Milhous WK. Neurotoxicity and efficacy of arteether related to its exposure times and exposure levels in rodents. *Am J Trop Med Hyg* **2002**;66:516–25.
53. Davies TME, Karunajeewa HA, Ilett KF. Artemisinin-based combination therapies for uncomplicated malaria. *Med J Aust* **2005**;182:181–5.
54. van Hensbroek MB, Onyiorah E, Jaffar S, et al. A trial of artemether or quinine in children with cerebral malaria. *N Engl J Med* **1996**;335:69–75.
55. Stepniewska K, Day N, Babiker A, et al. A meta-analysis using individual patient data of trials comparing artemether with quinine in the treatment of severe falciparum malaria. *Trans R Soc Trop Med Hyg* **2001**;95:637–50.
56. Dondorp A, Nosten F, Stepniewska K, Day N, White N; South East Asian Quinine Artesunate Malaria Trial (SEAQUAMAT) Group. Artesunate versus quinine for treatment of severe falciparum malaria: a randomised trial. *Lancet* **2005**;366:717–25.
57. Qian RS, Li ZL, Yu JL, Ma DJ. The immunologic and antiviral effect of qinghaosu. *J Tradit Chin Med* **1982**;2:271–6.
58. Kaptein SJ, Efferth T, Leis M, et al. The anti-malaria drug artesunate inhibits replication of cytomegalovirus in vitro and in vivo. *Antiviral Res* **2006**;69:60–9.
59. Efferth T, Marschall M, Wang X, et al. Antiviral activity of artesunate towards wild-type, recombinant, and ganciclovir-resistant human cytomegaloviruses. *J Mol Med* **2002**;80:233–42.
60. Shapira MY, Resnick IB, Chou S, et al. Artesunate as a potent antiviral agent in a patient with late drug-resistant cytomegalovirus infection after hematopoietic stem cell transplant. *Clin Infect Dis* **2008**;46:1455–7.
61. Marschall M, Freitag M, Weiler S, Sorg G, Stamminger T. Recombinant GFP-expressing human cytomegalovirus as a tool for screening of antiviral agents. *Antimicrob Agents Chemother* **2000**;44:1588–97.

62. Mocarski ES Jr, Shenk T, Pass RF. Cytomegalovirus. In: Fields BN, Knipe DM, Howley PM, eds. *Fields virology*. 5th ed. Philadelphia: Lippincott-Raven, **2007**:2701–72.
63. Yurochko AD, Kowalik TF, Huong SM, Huang ES. Human cytomegalovirus upregulates NF- $\kappa$ B activity by transactivating the NF- $\kappa$ B p105/p50 and p65 promoters. *J Virol* **1995**; *69*:5391–400.
64. Yurochko AD, Mayo MW, Poma EE, Baldwin AS Jr, Huang ES. Induction of the transcription factor Sp1 during human cytomegalovirus infection mediates upregulation of the p65 and p105/p50 NF- $\kappa$ B promoters. *J Virol* **1997**; *71*:4638–48.
65. Bork PM, Schmitz ML, Kuhnt M, Escher C, Heinrich M. Sesquiterpene lactone containing Mexican Indian medicinal plants and pure sesquiterpene lactones as potent inhibitors of transcription factor NF- $\kappa$ B. *FEBS Lett* **1997**; *402*:85–90.
66. Siedle B, Garcia-Pineres AJ, Murillo R, et al. Quantitative structure-activity relationship of sesquiterpene lactones as inhibitors of the transcription factor NF- $\kappa$ B. *J Med Chem* **2004**; *47*:6042–54.
67. Johnson RA, Wang X, Ma X-L, Huong S-M, Huang E-S. Human cytomegalovirus up-regulates the phosphatidylinositol 3-kinase (PI3-K) pathway: inhibition of PI3-K activity inhibits viral replication and virus-induced signaling. *J Virol* **2001**; *75*:6022–32.
68. Ghazal P, Lubon H, Fleckenstein B, Henninghausen L. Binding of transcription factors and creation of a large nucleoprotein complex on the human cytomegalovirus enhancer. *Proc Natl Acad Sci USA* **1987**; *84*:3658–62.
69. Prösch S, Wuttke R, Krüger DH, Volk HD. NF- $\kappa$ B—a potential therapeutic target for inhibition of human cytomegalovirus (re)activation? *Biol Chem* **2002**; *383*:1601–9.
70. Marchini A, Liu H, Zhu H. Human cytomegalovirus with IE-2 (UL122) deleted fails to express early lytic genes. *J Virol* **2001**; *75*:1870–8.
71. Eickhoff J, Hanke M, Stein-Gerlach M, et al. RICK activates an NF- $\kappa$ B dependent anti-HCMV response. *J Biol Chem* **2004**; *279*:9642–52.
72. Naesens L, Bonnafous P, Agut H, De Clercq E. Antiviral activity of diverse classes of broad-acting agents and natural compounds in HHV-6-infected lymphoblasts. *J Clin Virol* **2006**; *37*(Suppl 1):S69–75.
73. Fang CT, Nath N, Pielech M, Dodd RY. A modified technique for the detection of hepatitis B virus-specific DNA polymerase. *J Virol Methods* **1981**; *2*:349–56.
74. Lien J, Petcu DJ, Aldrich CE, Mason WS. Initiation of termination of duck hepatitis B virus DNA synthesis during virus maturation. *J Virol* **1987**; *61*:3832–40.
75. Wang GH, Seeger C. Novel mechanism for reverse transcription in hepatitis B viruses. *J Virol* **1993**; *67*:6507–12.
76. Bartholomeusz A, Locarnini S. Hepatitis B virus mutants and fulminant hepatitis B: fitness plus phenotype. *Hepatology* **2001**; *34*:432–5.
77. Iino S. Natural history of hepatitis B and C virus infections. *Oncology* **2002**; *62*(Suppl 1):18–23.
78. Beasley RP, Hwang LY. Overview on the epidemiology of hepatocellular carcinoma. In: Hollinger FB, Lemon SM, Margolis M, eds. *Viral hepatitis and liver disease*. Baltimore: Williams & Willkins, **1991**:532–5.
79. The EASL jury. EASL International Consensus Conference on Hepatitis B. *J Hepatol* **2003**; *38*:533–40.
80. Romero MR, Efferth T, Serrano MA, et al. Effect of artemisinin/artesunate as inhibitors of hepatitis B virus production in an “in vitro” replicative system. *Antiviral Res* **2005**; *68*:75–83.
81. Batty KT, Davis TM, Thu LT, Binh TQ, Anh T, Ilett KF. Selective high-performance liquid chromatographic determination of artesunate and alpha- and beta-dihydroartemisinin in patients with falciparum malaria. *J Chromatogr B Biomed Appl* **1996**; *677*:345–50.
82. Placidi L, Faraj A, Loi AG, et al. Antiviral activity and intracellular metabolism of bis(tButylSATE) phosphotriester of beta-l-2',3'-dideoxyadenosine, a potent inhibitor of HIV and HBV replication. *Antivir Chem Chemother* **2001**; *12*:99–108.
83. Tanikawa K. Pathogenesis and treatment of hepatitis C virus-related liver diseases. *Hepatobiliary Pancreat Dis Int* **2004**; *3*:17–20.
84. Buckwold VE, Beer BE, Donis RO. Bovine viral diarrhoea virus as a surrogate model of hepatitis C virus for the evaluation of antiviral agents. *Antiviral Res* **2003**; *60*:1–15.
85. Samuel CE. Antiviral actions of interferons. *Clin Microbiol Rev* **2001**; *14*:778–809.
86. Durantel D, Carrouee-Durantel S, Branza-Nichita N, Dwek RA, Zitzmann N. Effects of interferon, ribavirin, and iminosugar derivatives on cells persistently infected with noncytopathic bovine viral diarrhoea virus. *Antimicrob Agents Chemother* **2004**; *48*:497–504.
87. Lau JY, Tam RC, Liang TJ, Hong Z. Mechanism of action of ribavirin in the combination treatment of chronic HCV infection. *Hepatology* **2002**; *35*:1002–9.
88. Yang YZ, Little B, Meshnick SR. Alkylation of proteins by artemisinin: effects of heme, pH and drug structure. *Biochem Pharmacol* **1994**; *48*:569–73.
89. Romero MR, Serrano MA, Vallejo M, et al. Antiviral effect of artemisinin from *Artemisia annua* against a model member of the *Flaviviridae* family, the bovine viral diarrhoea virus (BVDV). *Planta Med* **2006**; *72*:1169–74.
90. Tam RC, Lau JY, Hong Z. Mechanisms of action of ribavirin in antiviral therapies. *Antivir Chem Chemother* **2001**; *12*:261–72.
91. Paeshuyse J, Coelmont L, Vliegen I, et al. Hemin potentiates the anti-hepatitis C virus activity of the antimalarial drug artemisinin. *Biochem Biophys Res Commun* **2006**; *348*:139–44.
92. Fillebeen C, Rivas-Estilla AM, Bisailon M, et al. Iron inactivates the RNA polymerase NS5B and suppresses subgenomic replication of hepatitis C virus. *J Biol Chem* **2005**; *280*:9049–57.
93. Birku Y, Mekonnen E, Bjorkman A, Wolday D. Delayed clearance of *Plasmodium falciparum* in patients with human immunodeficiency virus co-infection treated with artemisinin. *Ethiop Med J* **2002**; *40*:17–26.
94. Tang HQ, Hu J, Yang L, Tan RX. Terpenoids and flavonoids from *Artemisia* species. *Planta Med* **2000**; *66*:391–3.